Introduction

Gastric and esophageal cancers are the 3rd and 5th highest causes of cancer mortality worldwide, respectively, resulting in a total of 1.4 million deaths per year (1). In the United States, ~43,000 new cases and ~25,000 deaths from gastro-esophageal cancer are estimated to occur in 2016 (2). Most gastric cancers (GCs) are associated with Helicobacter Pylori (H. Pylori) and Epstein-Barr virus (EBV) infection, with chronic infection from H. Pylori causing about 90% of new cases of noncardiac GC worldwide (3-5). A small percentage of GCs arise from germline mutation in E-Cadherin.
(CDH1), which is associated with tumors displaying diffuse-type histology (6). Esophageal squamous cell carcinomas (ESCC) comprise 90% of esophageal cancers (ECs) in high-risk areas (including northern Iran, central Asia, and north-central China) and are thought to be related to poor nutritional status, low intake of vegetable and fruits, and high-temperature beverage drinking (1,7-10). Esophageal adenocarcinoma (EAC) is the more prevalent esophageal tumor in the U.S. and Europe, where its incidence has increased significantly in the last 30 years in parallel with obesity and gastro-esophageal reflex disease (11).

The need for identifying effective novel therapies is highlighted by the poor prognosis of patients with advanced gastro-esophageal cancer (survival ~12 months). Multiple randomized trials have shown negative results for agents targeting EGFR, VEGF, MTOR, and hedgehog. To date, only two biologic therapies have been shown to improve overall survival in these patients: trastuzumab, a monoclonal antibody (mAb) targeting human epidermal growth factor receptor 2 (HER2, ERBB2), and ramucirumab, a mAb targeting vascular endothelial growth factor receptor 2 (VEGFR2) (12-17).

Recent studies indicate that immune-modulating therapies may have efficacy in these tumors. A major question is whether molecular subsets can be identified in which these new therapies have increased efficacy. In this review article, we will discuss the available evidence for PD-1/PD-L1 blockade in gastro-esophageal cancers and the potential role of PD-L1 expression and microsatellite instability (MSI) as predictive biomarkers for PD-1/PD-L1 targeted treatment.

**PD-1/PD-L1 pathway**

The immune system can specifically identify and eliminate tumor cells on the basis of their expression of tumor-specific antigens or molecules induced by cellular stress (18). In this process, known as tumor immune surveillance or immunoeediting, the immune system identifies cancerous and/or precancerous cells and eliminates them before they can cause harm. However, if elimination is incomplete, a temporary state of equilibrium can develop between the immune system and the developing tumor. During this period it is envisaged that tumor cells either remain dormant or continue to evolve, accumulating further changes (such as DNA mutations or changes in gene expression) that can modulate the tumor-specific antigens and stress-induced antigens that they express. If the immune response still fails to completely eliminate the tumor, tumor cell variants are selected that are able to resist, avoid, or suppress the antitumor immune response, leading to immune escape.

Tumor infiltration by T cells, particularly cytotoxic T lymphocytes (CTLs), is part of the adaptive antitumor immune response and is believed to represent the elimination or equilibrium phase of immunoeediting. The presence of tumor infiltrating lymphocytes (TILs) has been associated with an improved prognosis for a number of different tumor types, including colorectal and GCs (19,20). CTL function is closely regulated by the tumor microenvironment, which consists of cancer cells, inflammatory cells, stromal cells and cytokines (21). The immunosuppressive network in the tumor microenvironment usually drives the CTL/TIL into “exhaustion”, a state of T-cell dysfunction marked by a unique molecular signature and that results in decreased cytokine expression and effector function (22).

Programmed cell death protein 1 (PD-1) pathway is considered an important inhibitory mechanism regulating T-cell exhaustion. PD-1 is expressed on the surface of T cells, B cells, monocytes and nature killer cells (23,24), including in TILs. PD-1 has two main ligands, PD-L1 and PD-L2. PD-L1 is expressed on T cells, B cells, dendritic cells, macrophages, mesenchymal stem cells and bone marrow-derived mast cells, and some non-hematopoietic cells. PD-L2 is expressed on dendritic cells, macrophages, bone marrow-derived mast cells, and resting peritoneal B1 cells (23,24). High expression of PD-L1 and PD-L2 has been detected on tumor cells (23,25). T cell functions are inhibited upon interaction between PD-1 and its ligand PD-L1 and/or PD-L2, which leads to T-cell exhaustion and is proposed as one mechanism underlying a tumor's ability to evade immune surveillance (26).

**PD-L1 expression in gastro-esophageal tumors**

Table 1 shows the results of studies which examined PD-L1 expression in human gastro-esophageal cancer tissues. PD-L1 has been detected in the tumor microenvironment of GCs including tumor cells, stromal and immune cells. The largest gastro-esophageal cancer cohort reported, to date, examined PD-L1 expression in 465 German gastric/gastro-esophageal junction (GEJ) cancer cases by immunohistochemistry (IHC) (antibody E1L3N clone, Cell Signaling) (27). The following expression patterns were found:

- PD-L1 expression on tumor cell membranes was detected in 30.1% of cases. The percentage of stained tumor cells was generally low [in 90% of the cases,
### Table 1: Frequency and prognostic value of PD-L1 expression in gastroesophageal cancer patients not treated with immune therapy

<table>
<thead>
<tr>
<th>Tumor type</th>
<th>N</th>
<th>Country</th>
<th>PD-L1 diagnostic antibody</th>
<th>PD-L1 expression</th>
<th>Immune/inflammatory cells in tumor microenvironment</th>
<th>Association of PD-L1 expression with clinical outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Gastric AC</td>
<td>465</td>
<td>Germany (27)</td>
<td>Eib3N clone signaling</td>
<td>IRS &gt;2(^a)</td>
<td>≥10%</td>
<td>35 Better survival(^e)</td>
</tr>
<tr>
<td>Gastric AC</td>
<td>398</td>
<td>China (28)</td>
<td>-</td>
<td>≥5%</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Esophageal AC</td>
<td>345</td>
<td>USA (29)</td>
<td>clone 405.9A11</td>
<td>≥5%</td>
<td>any</td>
<td>82% cases were PD-L2 positive</td>
</tr>
<tr>
<td>Gastric AC</td>
<td>180</td>
<td>Japan (30)</td>
<td>-</td>
<td>≥5%</td>
<td>-</td>
<td>Poor prognosis</td>
</tr>
<tr>
<td>Gastric AC</td>
<td>127</td>
<td>Germany (31)</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Poor prognosis</td>
</tr>
<tr>
<td>Gastric AC</td>
<td>111</td>
<td>China (32)</td>
<td>Abcam</td>
<td>10%</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Gastric AC</td>
<td>102</td>
<td>USA (29)</td>
<td>2H11</td>
<td>-</td>
<td>-</td>
<td>Advance stage and poor overall survival</td>
</tr>
<tr>
<td>Gastric/GEJ AC</td>
<td>34</td>
<td>USA (34)</td>
<td>5H1</td>
<td>&gt;5%</td>
<td>≥1%</td>
<td>45 -</td>
</tr>
<tr>
<td>Gastric AC</td>
<td>19</td>
<td>Japan (35)</td>
<td>-</td>
<td>≥1%</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Esophageal squamous cell cancer</td>
<td>41</td>
<td>Japan (36)</td>
<td>MIH1</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

\(^a\), IRS was determined for each sample (range, 0–7) and was defined as the sum of two parameters: the % of immunoreactive cells [0 (negative), 1 (≤1%), 2 (2–10%), 3 (11–50%), 4 (>50%)] and staining intensity [0, 1 (weak), 2 (moderate), 3 (strong)]; \(^b\), high PD-L1 expression was associated with male gender, EBV-positivity, microsatellite instability, proximal location within the stomach, HER2-positivity, PIK3CA-mutation; \(^c\), not reported. Abbreviations: AC, adenocarcinoma; GEJ, gastroesophageal junction; IRS, immunoreactivity score; NR, not reported; TIL, tumor infiltrating lymphocyte; TAM, tumor associated macrophage.
In the study, PD-L1 expression was observed neither in tumor nor in stroma cells. PD-1 diffusely distributed TILs were present in 53.8% of cases. PD-1 expression in TILs was significantly correlated with PD-L1 expression in tumor, stroma and immune cells.

PD-L1 tumor cell expression was detected more often in men and in gastric/GEJ cancers that were EBV-positive, MSI, proximally located, HER2-positive, or PIK3CA-mutated. The association between PD-L1 expression and a covariate was strongest for EBV, with 18 of 20 EBV-positive cases showing PD-L1 expression in tumor cells and 14 of 20 EBV-positive cases showing PD-L1 expression in immune cells.

Likewise, a study of PD-L1 expression and immune infiltration in GCs from Asia (N=398 cases) found that 14% of cases were PD-L1 positive in tumor cells. A sample was considered for PDL1 expression if 5% or more of tumor cells showed membrane positivity. There was correlation between PD-L1 expression and density of TILs.

In both studies, PD-L1 tumor-cell positivity had an association with favorable overall survival that was either statistically significantly (27) or trended toward significance (28). However, other investigations in gastro-esophageal cancer have reported an adverse association between PD-L1 expression and survival (30-33,35,36).

The frequency of PD-L1 expression in EAC was recently reported to be lower than what has been reported to date in GC. PD-1, PD-L1, and PD-L2 expression was analyzed on a tissue microarray (TMA) containing EAC samples from 345 patients. Surprisingly, only 1.7% of cases had PD-L1 positive tumor cells, and only 18% of cases had PD-L1+ staining in inflammatory cells (mostly macrophages). An evaluation of whole-tumor sections from a subset of 45 tumors revealed that the frequency of PD-L1 positive immune cells was somewhat higher [35.6% (16/45) of cases, of which 7/16 were not identified on the TMA], although still lower than that reported in GC. Interestingly, Barrett’s-associated EAC has been reported to have a low frequency of MSI and EBV (see below) (37,38).

On the other hand, 81.6% of EACs were found to be PD-L2 positive in at least one core. While PD-L2 and PD-L1 expression were not mutually exclusive, tumors with PD-L2 expression in all evaluated cores were less likely to possess PD-L1+ immune cells. Both PD-L2+ and PD-L1+ tumors had a higher average number of PD-1+ TILs compared to tumors without PD-L2 or PD-L1 expression. In 15.5% (53/343) of EACs, no PD-L2+, PD-L1+, or PD-1+ cells were observed.

The investigators suggested that the high frequency of PD-L2 expression, in the absence of PD-L1 co-expression, may be due to the fact that EACs develop in a background of chronic inflammation and typically emerge from Barrett’s esophagus (BE), a tissue with a documented Th2-skewed inflammatory state with increased IL4/IL13 expression (39,40). In macrophages and dendritic cells PD-L2 transcription is regulated by IL4/IL13/STAT6 signaling (41,42), raising the possibility that PD-L2 epithelial expression in EAC and BE may result from IL4/IL13 expression (29,43). The therapeutic role of inhibiting PD-L2 is controversial. While some studies show an inhibitory role for PD-L2 (44,45), others suggest that PD-L2 can stimulate T-cell proliferation (46) via a PD-1-receptor independent mechanism, potentially involving a distinct PD-L2 binding partner. Further evaluation of PD-L2 expression in independent EAC cohorts is warranted, including its potential role in predicting response to PD-1 blockade.

The varying rates of PD-L1 positivity and its prognostic value may be due to different patient populations (e.g., race, disease stage), different antibodies, and different cut-points. However, taken together, these studies indicate that PD-1/ PD-L1 expression occurs in a subset of tumors, suggesting a molecular target exists for therapeutic inhibition of the PD-1/PD-L1 pathway. In addition, these data suggest that the immune microenvironment of EAC may differ from gastric/GEJ adenocarcinomas, and should be an area of further investigation.
Anti-PD-1/PD-L1 therapy in gastro-esophageal tumors

PD-1/PD-L1 blockade has recently been shown to be a promising treatment in a variety of tumor types (47-50). The PD-1 inhibitors, pembrolizumab and nivolumab, are both immunoglobulin G4 (IgG4) antibodies, which bind to PD-1 to disrupt the interaction between PD-1 and its ligands and thereby impede inhibitory signals in T cells (51,52). Pembrolizumab is FDA-Approved for the treatment of unresectable or metastatic melanoma and for PD-L1 positive metastatic non-small cell lung cancer (NSCLC) (47,53-55). Nivolumab is FDA-approved for the treatment as a single agent or in combination with ipilimumab for the treatment of unresectable or metastatic melanoma, metastatic NSCLC that progresses on or after platinum based chemotherapy, and advanced renal cell carcinoma after anti-angiogenic therapy (49,50,56-62).

Table 2 shows the results from trials, to date, which examined anti-PD-1/PD-L1 therapy in gastro-esophageal cancer patients. The first reported evaluation of a PD-1/PD-L1 inhibitor in esophageal or GC examined pembrolizumab in PD-L1-positive GC patients. In this study (KEYNOTE-012), 162 patients with advanced gastric/GEJ cancer (recurrent or metastatic) were screened for PD-L1 expression by 22C3 antibody IHC staining. Only patients with distinct stromal or ≥1% tumor nest cell PD-L1 staining were eligible. A total of 65 patients were considered PD-L1 positive, of which 39 were enrolled onto the trial and treated with pembrolizumab. A majority of patients [66.7% (16/39)] received more than one prior treatment. The overall response rate (ORR) was 22% by central review. After a median follow up of 8.8 months, the 6-month progression free survival (PFS) rate was 24% and 6-month overall survival (OS) rate was 69%. Median time to response was 8 weeks with a median response duration (RD) of 24 weeks. The drug appeared to be well tolerated. Four patients experienced grade 3–5 drug-related adverse events, including fatigue, decreased appetite, peripheral sensory neuropathy, hypoxia and pneumonitis. One patient died of hypoxia (64).

As shown in Table 3, the ORR observed for pembrolizumab monotherapy compares favorably to single-agent response rates observed in randomized trials for ramucirumab in the second-line setting (ORR <4%, duration of therapy 8 weeks; REGARD) or regorafenib in the first-/second-line setting (ORR 3%, duration of therapy 8 weeks; INTEGRATE). However, response rates were higher in the RAINBOW study for paclitaxel arm (RR ~17%) and paclitaxel-ramucirumab arm (RR ~28%). In addition, the median PFS and OS observed for pembrolizumab is comparable to those reported in second-line trials studying these other therapies (15,17). While cross-trial comparisons must be viewed with caution, the non-randomized data from KEYNOTE-012 provide proof-of-concept that anti-PD-1 therapy may have activity in advanced GC and warrants further study.

Nivolumab has also demonstrated single-agent activity. In CheckMate-032, patients with solid tumors were treated with nivolumab with or without ipilimumab. Preliminary results from a subset of patients with advanced gastric or GEJ cancer of any PD-L1 status (n=59) who were treated with nivolumab monotherapy demonstrated an ORR of 12% (1 CR, 6 PRs) and a median RD of 7.1 months (66).

Table 2 also shows available data from studies examining the PD-L1 antibodies avelumab (MSB0010718C), atezolizumab (MPDL3280A), and durvalumab (MEDI4716). In general, RRs in gastro-esophageal cancer patients for these antibodies have ranged from 10% to 30% (35,63,65,67,70-73).

Given the variability of response to PD-1 blockade, research attention has concurrently focused on identifying biomarkers of response so as to select patients who are most likely to benefit. The candidate biomarker studied most extensively is PD-L1 expression in trials utilizing PD-1 blockade.

PD-L1 expression as a predictive marker for PD-1/PD-L1 blockade

At least five large studies have examined PD-L1 expression as a predictive biomarker of anti-PD-1 therapy in patients with advanced carcinoma. A phase 1 study of pembrolizumab monotherapy in 495 NSCLC patients (Keynote-001, NCT10295827), consisting of both squamous and non-squamous histology, showed a correlation between PD-L1 expression and ORR (52). Using the anti-PD-L1 antibody clone 22C3 (Merck), PD-L1 expression was analyzed in the membrane of tumor cells and in intercalated mononuclear inflammatory cells within tumor nests and stroma adjacent to tumor nests. After the study was initiated, membranous PD-L1 expression in at least 50% of tumor cells (proportion score, ≥50%) was selected as the cut-off on the basis of the ease of use and receiver-operating-characteristic (ROC) analysis. ORRs were 45%, 17% and 11% in the overall population with PD-L1 proportion scores of ≥50%, 1–49% and ≤1%, respectively. Median PFS among patients with a PD-L1 proportion score of at least 50% was 6.3 months (95% CI, 2.9 to 12.5), and was shorter among patients with a...
### Table 2 Results of PD-1/PD-L1 blockade in gastroesophageal cancer patients and predictive value of PD-L1 expression

<table>
<thead>
<tr>
<th>Study medication</th>
<th>Target</th>
<th>Phase</th>
<th>Cancer types</th>
<th>N</th>
<th>Clinical Trials Identifier (study status)</th>
<th>PD-L1 positive rate (%)</th>
<th>Response rate (data refer to gastroesophageal cancer patients unless noted otherwise) (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Overall</td>
</tr>
<tr>
<td>Atezolizumab</td>
<td>PD-L1</td>
<td>1</td>
<td>locally advanced or metastatic solid tumors</td>
<td>171&lt;sup&gt;a&lt;/sup&gt;</td>
<td>NCT01375842 (recruiting)</td>
<td>–&lt;sup&gt;d&lt;/sup&gt;</td>
<td>21&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pembrolizumab</td>
<td>PD-1</td>
<td>1</td>
<td>Advanced solid tumors (includes PD-L1+ GC)</td>
<td>39 (GC)</td>
<td>NCT01848834 (KEYNOTE-012) (finished recruiting)</td>
<td>65</td>
<td>22&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Pembrolizumab</td>
<td>PD-1</td>
<td>1</td>
<td>Advanced solid tumors</td>
<td>23 (EC)</td>
<td>NCT02054806 (KEYNOTE-028) (finished recruiting)</td>
<td>–&lt;sup&gt;d&lt;/sup&gt;</td>
<td>30</td>
</tr>
<tr>
<td>Nivolumab</td>
<td>PD-1</td>
<td>1/2</td>
<td>Advanced or metastatic solid tumors</td>
<td>59 (GC/GEJ)</td>
<td>NCT01928394 (CheckMate-032) (recruiting)</td>
<td>38</td>
<td>12</td>
</tr>
<tr>
<td>Avelumab</td>
<td>PD-L1</td>
<td>1</td>
<td>locally advanced or metastatic solid tumors</td>
<td>20 (GC/GEJ)</td>
<td>NCT01943461 (Japanese JAVELIN) (recruiting)</td>
<td>26</td>
<td>15</td>
</tr>
<tr>
<td>Avelumab</td>
<td>PD-L1</td>
<td>1</td>
<td>locally advanced or metastatic solid tumors</td>
<td>75 (GC/GEJ)</td>
<td>NCT01772004 (Korean JAVELIN) (recruiting)</td>
<td>–&lt;sup&gt;d&lt;/sup&gt;</td>
<td>9</td>
</tr>
<tr>
<td>Nivolumab</td>
<td>PD-1</td>
<td>2</td>
<td>Advanced esophageal cancer</td>
<td>65</td>
<td>NR (finished recruiting)</td>
<td>–&lt;sup&gt;d&lt;/sup&gt;</td>
<td>17</td>
</tr>
</tbody>
</table>

<sup>a</sup>, precise number of esophagogastric patients has not been reported; <sup>b</sup>, only PD-L1 positive cases received treatment; <sup>c</sup>, RRs in overall population, including non-gastric/GEJ cancers; <sup>d</sup>, not reported. Abbreviations: EC, esophageal cancer; GC, gastric cancer; GEJ, gastroesophageal junction; ORR, overall response rate; RR, response rate.
Table 3 Efficacy results in advanced gastroesophageal cancer from trials in the second-line or higher setting

<table>
<thead>
<tr>
<th>Study</th>
<th>Treatment arm</th>
<th>ORR (%)</th>
<th>PFS rate at 6 months (%)</th>
<th>Median PFS (months)</th>
<th>OS rate at 6 months (%)</th>
<th>Median OS (months)</th>
<th>DOR (months)</th>
</tr>
</thead>
<tbody>
<tr>
<td>RAINBOW (17)</td>
<td>Ramucirumab + Paclitaxel</td>
<td>28</td>
<td>36</td>
<td>4.4</td>
<td>72</td>
<td>9.6</td>
<td>-a</td>
</tr>
<tr>
<td>KEYNOTE-012 (64)</td>
<td>Pembrolizumab</td>
<td>22</td>
<td>24</td>
<td>69%</td>
<td>69</td>
<td>Not reached</td>
<td>6</td>
</tr>
<tr>
<td>RAINBOW (17)</td>
<td>Paclitaxel</td>
<td>~17</td>
<td>17</td>
<td>2.9</td>
<td>57</td>
<td>7.2</td>
<td>-a</td>
</tr>
<tr>
<td>CHECKMATE-032 (66)</td>
<td>Nivolumab</td>
<td>12</td>
<td>-a</td>
<td>-a</td>
<td>49</td>
<td>5</td>
<td>7.1</td>
</tr>
<tr>
<td>REGARD (15)</td>
<td>Ramucirumab</td>
<td>&lt;4</td>
<td>&lt;20</td>
<td>2.1</td>
<td>75</td>
<td>5.2</td>
<td>-a</td>
</tr>
<tr>
<td>INTEGRATE (69)</td>
<td>Regorafenib</td>
<td>3</td>
<td>&lt;20</td>
<td>2.6</td>
<td>75</td>
<td>5.8</td>
<td>-</td>
</tr>
</tbody>
</table>

a, not reported. Abbreviations: ORR, overall response rate; PFS, progression free survival; OS, overall survival, DOR, duration of response.

The predictive value of PD-L1 expression was also analyzed in squamous vs. non-squamous NSCLC populations separately in two studies. In both studies, patients with advanced NSCLC were randomized to nivolumab or docetaxel in the second-line setting. Identical methodology was used to assess PD-L1 expression in both studies. PD-L1 expression levels were analyzed in pre-treatment (archival or recent) tumor biopsy specimens using a validated automated IHC assay (Epitomics, clone 28-8). Samples were categorized as positive when staining of the tumor-cell membrane (at any intensity) was observed in a pre-specified percentage of cells (1%, 5%, or 10%) in a section that included at least 100 evaluable tumor cells. Despite similar methodology, the conclusions of one study differed from the other.

The non-squamous lung cancer trial (CheckMate-057, N=582) was a randomized, open-label, international phase 3 study of patients with non-squamous NSCLC [93% had adenocarcinoma (AC)] that had progressed during or after platinum-based doublet chemotherapy. Patients were treated with nivolumab (3 mg/kg every 2 weeks) or docetaxel (75 mg/m² every 3 weeks). The clinical study met its primary endpoint of OS, with a median OS of 12.2 months in the nivolumab group and 9.4 months in the docetaxel group (P=0.002). The response rate was 19% with nivolumab versus 12% with docetaxel (P=0.02). Although median PFS did not favor nivolumab over docetaxel (2.3 vs. 4.2 months, respectively), the 1-year PFS rate was higher with nivolumab than with docetaxel (19% and 8%, respectively).

In this trial, 78% (455/582) of randomized patients had quantifiable PD-L1 expression. The rate of PD-L1 positivity was 54% (123/228) at the 1% cut-point, 40% (181/455) at the 5% cut-point, and 36% (165/455) at the 10% cut-point. Nivolumab was associated with greater efficacy than docetaxel across all end points (RR, PFS, and OS) in subgroups defined according to pre-specified levels of tumor-membrane expression (≥1%, ≥5%, and ≥10%) of PD-L1. As an example, at the 10% cut-point for PD-L1 expression, the RR from nivolumab in PD-L1 positive vs. negative tumors was 37% vs. 11%, whereas docetaxel-treated patients had a RR of ~14% regardless of PD-L1 status. For PFS, among PD-L1 positive tumors, nivolumab-treated patients had a median PFS of 5.0 months, as compared to 3.7 months among docetaxel-treated patients [HR 0.52 (0.37, 0.75)]. By contrast, there was no PFS benefit for nivolumab among PD-L1 negative tumors [median PFS 2.1 months with nivolumab vs. 4.2 months with docetaxel; HR 1.24 (0.96, 1.61)]. Results were similar for OS, as follows: Among PD-L1 positive tumors, nivolumab-treated patients had a median OS of 19.9 months, as compared to 8.0 months among docetaxel-treated patients [HR 0.40 (0.27, 0.58)]. By contrast, there was no OS benefit for nivolumab among PD-L1 negative tumors [median PFS 9.9 months with nivolumab vs. 10.3 months with docetaxel; HR 0.96 (0.74, 1.25)]. Importantly, interaction P values were statistically significant for all PD-L1 expression cut-point levels and clinical endpoints (with the exception of the 1% cut-point for OS), strongly indicating
PD-L1 expression was predictive of nivolumab benefit. A caveat is that randomization was not stratified by PD-L1 expression—i.e., the PD-L1 subgroup analysis disrupted randomization and thus is not definitive. The authors indicated that the improved safety profile and durability of responses to nivolumab suggest that it might be a reasonable option for patients regardless of PD-L1 expression.

By contrast, PD-L1 expression was not found to be predictive in a smaller trial of squamous lung cancers (checkmate-017, N=272). In this trial, patients with molecularly unselected stage IIIB or IV cancer who had disease progression after one prior platinum-containing regimen were randomized to nivolumab vs. docetaxel. An OS benefit favoring nivolumab over chemotherapy alone was demonstrated (median OS of 9.2 vs. 6.0 months, respectively). A total of 83\% (225/272) of randomized patients had quantifiable PD-L1 expression. The rate of PD-L1 positivity was 53\% (119/225) at the 1\% cut-point, 36\% (81/225) at the 5\% cutpoint, and 31\% (69/225) at the 10\% cut-point. The authors reported that PD-L1 expression was neither prognostic nor predictive of RR, PFS, or OS across all pre-specified expression levels of PD-L1 (1\%, 5\%, and 10\%). While the survival curves suggested a slightly greater benefit from nivolumab for PD-L1 positive tumors as compared to PD-L1 negative tumors [e.g., HR for PFS 0.54 (95\% CI 0.32, 0.90)] for the PD-L1 positive group at the 5\% cut-point vs. HR 0.75 [(0.52, 1.1) for the PD-L1 negative group], nivolumab prolonged OS as compared to docetaxel in patients with PD-L1 negative tumors—indicating that PD-L1 tumor expression was not a strong predictive marker (49).

It is possible that the difference in results for PD-L1 expression as a predictive marker between the two NSCLC studies may be due to a difference in the immune milieu between squamous vs. non-squamous NSCLCs, which is an area that should be studied further. However, dynamic PD-L1 expression related to the tumor microenvironment (74) and responses observed in patients with low PD-L1 expression levels have raised questions on whether PD-L1 expression is an ideal marker for PD-1 treatment.

In support of this concept, PD-L1 expression was found not to be predictive in a phase 3 trial (CheckMate-025; N=821) of previously treated advanced clear-cell renal-cell carcinoma (RCC). In this trial, patients randomized to receive nivolumab had an OS of 25 months, as compared to 19.6 months for patients in the everolimus arm. PD-L1 expression was not predictive for response across different expression levels. Median OS in the nivolumab arm were 21.8 months and 27.4 months for tumors with PD-L1 expression levels of ≥1\% or <1\%, respectively. Results for OS were similarly null using a 5\% cutpoint of PD-L1 expression (50).

More recent data have indicated that PD-L1 expression was predictive of OS benefit in NSCLC patients treated with an anti-PD-L1 Ab. Advanced NSCLC patients (N=287; both non-squamous and squamous) were randomized to atezolizumab (an anti-PD-L1 Ab) vs. docetaxel in the second-line setting (75). This phase 2 trial met its primary endpoint of OS (HR 0.73 favoring atezolizumab). Moreover, the benefit in OS was limited to patients whose tumors had any PD-L1 expression in either the tumor or in tumor-infiltrating immune cells [HR 0.59 (95\% CI 0.40–0.85)]. By contrast, patients with no PD-L1 expression had no benefit in OS [HR 1.04 (95\% CI 0.62–1.75)]. A strength of this study, while not large, is that patients were stratified by PD-L1 expression status. PD-L1 expression status also predicted for improved RR and PFS, although less strongly. This therapeutic antibody and candidate predictive biomarkers are currently being examined in larger NSCLC cohorts.

In advanced gastric/GEJ cancer, data are more limited regarding the predictive value of PD-L1 expression. In KEYNOTE-012 (primary results described above), where pembrolizumab monotherapy was administered only to patients with PD-L1 expressing tumors, a trend toward an association between higher levels of PDL1 expression and ORR, PFS, and OS was observed (1-sided P=0.10, 0.16, and 0.12, respectively) (14). Further detail has not been reported. Given the small sample size, and because PD-L1 non-expressing tumors were excluded from KEYNOTE-012, it is currently difficult to say whether PD-L1 expression correlates with pembrolizumab efficacy. The ongoing KEYNOTE-059 study is enrolling patients with advanced gastric/GEJ cancer in both the first- and third-line settings, including both PD-L1 positive and negative tumors and thus may be able to address whether PD-L1 expression is a predictive marker for pembrolizumab. In addition, this question can be further examined in two new phase 3 trials (Table 4): a second-line trial comparing pembrolizumab with paclitaxel (KEYNOTE-061; NCT02370498), which includes tumors of any PD-L1 status; and a new first-line trial comparing pembrolizumab vs. cisplatin/5-FU vs. cisplatin/5-FU plus pembrolizumab, which includes only PD-L1 overexpressing tumors (KEYNOTE-062).

For nivolumab monotherapy in gastric/GEJ cancer, the CheckMate-032 study (primary results described above)
Table 4 Ongoing trials examining PD-1/PD-L1 blockade in gastroesophageal cancers\textsuperscript{a}

<table>
<thead>
<tr>
<th>Clinical Trial Identifier</th>
<th>Study medication</th>
<th>Cancer type</th>
<th>Phase</th>
<th>Primary endpoints</th>
<th>Comments (72,73,76-80)</th>
</tr>
</thead>
<tbody>
<tr>
<td>NCT02678182</td>
<td>Durvalumab vs. capcitabine vs. trastuzumab vs. surveillance</td>
<td>EAC/GC/GEJ</td>
<td>2</td>
<td>PFS</td>
<td>Maintenance treatment. Durvalumab is an anti-PD-L1 inhibitor</td>
</tr>
<tr>
<td>NCT02625610 (JAVELIN Gastric 100)</td>
<td>Avelumab vs. oxaliplatin-FP</td>
<td>GC/GEJ</td>
<td>3</td>
<td>OS, PFS</td>
<td>1st-line</td>
</tr>
<tr>
<td>NCT02370498 (KEYNOTE-061)</td>
<td>Pembrolizumab vs. paclitaxel</td>
<td>GC/GEJ</td>
<td>3</td>
<td>PFS, OS in PD-L1 positive participants</td>
<td>2nd-line</td>
</tr>
<tr>
<td>NCT02564263 (KEYNOTE-181)</td>
<td>Pembrolizumab vs. chemo</td>
<td>EC/GEJ</td>
<td>3</td>
<td>PFS, OS</td>
<td>After first-line: physician's choice of chemo includes single agent docetaxel, paclitaxel, or irinotecan</td>
</tr>
<tr>
<td>NCT02267343</td>
<td>Nivolumab vs. placebo</td>
<td>GC</td>
<td>3</td>
<td>OS</td>
<td>For patient who failed standard chemotherapy</td>
</tr>
<tr>
<td>NCT02569242</td>
<td>Nivolumab vs. docetaxel/paclitaxel</td>
<td>EC</td>
<td>3</td>
<td>OS</td>
<td>2nd-line: patients will be divided into 4 subgroups based on gene expression profiling and TP53 status</td>
</tr>
<tr>
<td>NCT02625623 (JAVELIN Gastric 300)</td>
<td>Avelumab vs. chemo</td>
<td>GC/GEJ</td>
<td>3</td>
<td>OS</td>
<td>3rd line</td>
</tr>
<tr>
<td>NCT02589496</td>
<td>Pembrolizumab</td>
<td>GC/GEJ</td>
<td>2</td>
<td>RR</td>
<td>1st, 2nd, 3rd line, maintenance treatment</td>
</tr>
<tr>
<td>KEYNOTE-059</td>
<td>Pembrolizumab</td>
<td>GC/GEJ</td>
<td>2</td>
<td>ORR</td>
<td>3rd- and 1st-line (first-line includes combination with cisplatin/FP)</td>
</tr>
<tr>
<td>NCT02559687 (KEYNOTE-190)</td>
<td>Pembrolizumab</td>
<td>EC/GEJ</td>
<td>2</td>
<td>ORR</td>
<td>3rd line</td>
</tr>
<tr>
<td>NCT02710396</td>
<td>Pembrolizumab</td>
<td>ESCC and other tumors</td>
<td>2</td>
<td>ORR</td>
<td>3rd line</td>
</tr>
<tr>
<td>NCT01772004 (JAVELIN Solid Tumor)</td>
<td>Avelumab</td>
<td>GC/GEJ and other solid tumors</td>
<td>1</td>
<td>DLTs, ORR</td>
<td>3rd line, maintenance treatment</td>
</tr>
<tr>
<td>Combined with other therapy in advanced disease</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NCT02494583 (KEYNOTE-062)</td>
<td>Pembrolizumab vs. cisplatin/FP with pembrolizumab vs. cisplatin/FP</td>
<td>GC/GEJ</td>
<td>3</td>
<td>PFS, OS</td>
<td>1st-line</td>
</tr>
<tr>
<td>NCT02340975</td>
<td>Durvalumab with tremelimumab vs. tremelimumab</td>
<td>GC/GEJ</td>
<td>1/2</td>
<td>AEs, DLT, ORR, PFS</td>
<td>Tremelimumab is a human anti-CTLA-4 monoclonal antibody</td>
</tr>
<tr>
<td>NCT02658214</td>
<td>Durvalumab with tremelimumab</td>
<td>GC/GEJ</td>
<td>1</td>
<td>AEs, lab findings</td>
<td>1st-line</td>
</tr>
<tr>
<td>NCT02734004</td>
<td>Durvalumab with olaparib</td>
<td>ATM-negative GC and other solid tumor</td>
<td>1/2</td>
<td>DCR, safety, tolerability</td>
<td>Olaparib is a Poly (adenosine diphosphate-ribose) polymerase (PARP) inhibitor</td>
</tr>
<tr>
<td>NCT02572687</td>
<td>Durvalumab with ramucirumab</td>
<td>GC/GEJ</td>
<td>1</td>
<td>DLTs</td>
<td>Epacadostat is an inhibitor of indoleamine 2,3-dioxygenase (IDO1)</td>
</tr>
<tr>
<td>NCT02318277</td>
<td>Durvalumab with epacadostat</td>
<td>GC/GEC and other solid tumor</td>
<td>1/2</td>
<td>DLTs, ORR</td>
<td></td>
</tr>
<tr>
<td>NCT02689284</td>
<td>Pembrolizumab and margetuximab</td>
<td>HER2-positive GC/GEJ</td>
<td>1/2</td>
<td>MTD, MAD of margetuximab, RD, ORR</td>
<td>Margetuximab is a HER2-targeted agent</td>
</tr>
</tbody>
</table>

\textsuperscript{a} Table 4 (continued)
Table 4 (continued)

<table>
<thead>
<tr>
<th>Clinical Trial Identifier</th>
<th>Study medication</th>
<th>Cancer type</th>
<th>Phase</th>
<th>Primary endpoints</th>
<th>Comments (72,73,76-80)</th>
</tr>
</thead>
<tbody>
<tr>
<td>NCT02452424</td>
<td>Pembrolizumab and PLX3397</td>
<td>Melanoma and other solid tumors (includes GC)</td>
<td>1/2</td>
<td>AEs</td>
<td>PLX3397 is designed to target the receptor for CSF1 (CSF1R)</td>
</tr>
<tr>
<td>NCT02318901 (PembroMab)</td>
<td>Pembrolizumab and monoclonal antibody</td>
<td>HER2-positive GC/GEJ</td>
<td>1/2</td>
<td>RP2D of Mab</td>
<td>Pembrolizumab and trastuzumab is one cohort</td>
</tr>
<tr>
<td>NCT02563548</td>
<td>Pembrolizumab and PEGPH20</td>
<td>NSCLC or GC</td>
<td>1b</td>
<td>DLT, ORR</td>
<td>PEGPH20 is a PEGylated form of recombinant human hyaluronidase (HA); PEGPH20 is designed to degrade HA</td>
</tr>
<tr>
<td>NCT01174121</td>
<td>Pembrolizumab with TILs</td>
<td>GC</td>
<td>2</td>
<td>Rate of tumor regression</td>
<td>TILs and pembrolizumab is one arm</td>
</tr>
<tr>
<td>NCT02443324</td>
<td>Pembrolizumab and ramucirumab</td>
<td>GC/GEJ</td>
<td>1</td>
<td>DLTs</td>
<td>GC/GEJ cancer is one of three tumor types under study</td>
</tr>
<tr>
<td>NCT02268825</td>
<td>Pembrolizumab with mFOLFOX</td>
<td>GI cancers (includes gastro-esophageal)</td>
<td>1/2</td>
<td>Safety</td>
<td>ESCC is excluded</td>
</tr>
<tr>
<td>NCT02642809</td>
<td>Pembrolizumab with local radiation</td>
<td>EC</td>
<td>0</td>
<td>Tolerability</td>
<td></td>
</tr>
<tr>
<td>NCT01928394</td>
<td>Nivolumab vs. nivolumab with Ipilimumab</td>
<td>Solid tumors (includes GC)</td>
<td>1/2</td>
<td>ORR</td>
<td>Chemotherapy is oxaliplatin with either Tegafur or capecitabine</td>
</tr>
<tr>
<td>NCT02746796</td>
<td>Nivolumab with chemotherapy vs. chemotherapy</td>
<td>GC/GEJ</td>
<td>2</td>
<td>ORR</td>
<td></td>
</tr>
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</table>

Curative-intent setting

<table>
<thead>
<tr>
<th>Clinical Trial Identifier</th>
<th>Study medication</th>
<th>Cancer type</th>
<th>Phase</th>
<th>Primary endpoints</th>
<th>Comments (72,73,76-80)</th>
</tr>
</thead>
<tbody>
<tr>
<td>NCT02743494 (CheckMate 577)</td>
<td>Adjuvant nivolumab vs. placebo</td>
<td>EC/GEJ</td>
<td>3</td>
<td>DFS, OS</td>
<td>Therapy is given after chemoradiation and surgery</td>
</tr>
<tr>
<td>NCT02488759 (CheckMate 358)</td>
<td>Neoadjuvant nivolumab</td>
<td>Virus-associated tumors (includes GC/GEJ)</td>
<td>1/2</td>
<td>AEs, SAEs, ORR, rate of surgery delay</td>
<td>This trial has neoadjuvant and metastatic cohorts</td>
</tr>
<tr>
<td>NCT02735239</td>
<td>Neoadjuvant durvalumab with chemo(radio) therapy</td>
<td>EC</td>
<td>1/2</td>
<td>AEs, DLTs, Lab findings</td>
<td>Expansion cohorts include durvalumab in combination with tremelimumab and chemotherapy and/or radiation.</td>
</tr>
<tr>
<td>NCT02520453</td>
<td>Adjuvant durvalumab</td>
<td>ESCC</td>
<td>2</td>
<td>DFS</td>
<td>Therapy is given after standard neoadjuvant chemoradiotherapy</td>
</tr>
<tr>
<td>NCT02639065</td>
<td>Adjuvant durvalumab</td>
<td>EC</td>
<td>2</td>
<td>RFS</td>
<td>Therapy is given after standard neoadjuvant chemoradiotherapy</td>
</tr>
<tr>
<td>NCT02730546</td>
<td>Neoadjuvant pembrolizumab with chemoradiotherapy</td>
<td>Locally advanced, resectable GEJ or gastric cardia</td>
<td>1/2</td>
<td>PathCR, PFS</td>
<td>Adjuvant pembrolizumab is also given</td>
</tr>
</tbody>
</table>

*, trials shown are those listed in clinicaltrials.gov as of 4/23/2016. Abbreviations: EC, esophageal cancer; ESCC, esophageal squamous cell cancer; FP, fluoropyrimidine; GC, gastric cancer; GEJ, gastroesophageal junction; MTD, maximum tolerated dose; MAD, maximum administered dose; RD, response duration; ORR, overall response rate; OS, overall survival; PFS, progression free survival; AEs, advance events; RP2D, recommended phase 2 dose; Mab, monoclonal antibody; DCR, disease control rate; ATM, ataxia telangiectasia mutated; NSCLC, non-small cell lung cancer; RFS, relapse free survival; irPFS, immune-response related assessment of progression free survival; irTTP, immune-response related assessment of time to tumor progression; irORR, immune-response related assessment of time to tumor progression; irOS, immune-response related assessment of overall survival; DLTs, dose limiting toxicities; DOR, duration of response; TTR, time to first response; PK, pharmacokinetics; QOL, quality of life; TEAEs, treatment emergent adverse events; HER2, human epidermal growth factor receptor 2; DFS, disease free survival; PathCR, pathological complete remission.
enrolled patients of any PD-L1 status (66). A sample was considered positive if PD-L1 expression was observed on the membrane of tumor cells. These criteria differ from those used in KEYNOTE-012, which considered expression in tumor or immune cell as positive. In CheckMate-032, two different cut-offs for PD-L1 expression were considered positive: 1% or more of tumor cells, and 5% or more. At the 1% cut-off, 38% (15/40) of cases were PD-L1 positive; and at the 5% cut-off, 15% (6/40) were PD-L1 positive. The ORR was numerically higher in PD-L1-positive vs. -negative tumors at both the 1% cut-off (27% in PD-L1 positive tumors vs. 12% in PD-L1 negative tumors) and the 5% cut-off (33% vs. 15%). This trend suggests that PD-L1 expression could be predictive for nivolumab efficacy and is consistent with the data regarding pembrolizumab monotherapy. While intriguing, these data require confirmation in larger cohorts.

Taken together, data from large clinical trial populations of advanced carcinoma (excluding melanoma) suggests that baseline PD-L1 expression in the tumor membrane may have some value in predicting efficacy from PD-1/PD-L1 blockade. However, the data are inconsistent across tumor types, and a potential benefit from checkpoint inhibition has not been ruled out in patients with PD-L1-negative tumors. Data from gastric/GEJ cancer are sparse and similarly suggest that PD-L1 expression may have some predictive value, but to date, has not ruled out benefit for patients whose tumors lack expression. The inconsistence of results may reflect the fact that PD-L1 and PD-1 expression are dynamic markers that change in relation to local cytokines and other factors, and they may be an imperfect surrogate for an “immunogenic” tumor microenvironment that has higher rates of functionally exhausted T cells infiltrating the tumor. In addition, the varying results between studies could be due to differences in antibodies used, definitions of positivity (cut-off percentage, expression in tumor cells vs. infiltrating lymphocytes, membranous vs. cytoplasmic expression, etc.) (81-84). Moving forward, eligibility for trials in gastro-esophageal cancer examining PD-1 blockade do not generally appear to be limited to a particular PD-L1 status, which will allow the ability to address the question of its predictive value more definitively.

**Mismatch repair (MMR) deficiency and hypermutation in gastro-esophageal tumors**

Deficiencies in the DNA MMR system cause errors during DNA replication, which in turn give rise to MSI. Microsatellites, which are simple repeat sequences of 1 to 6 base pairs (also known as short tandem repeats), are particularly prone to DNA replication errors (85). MSI manifests as small increases or decreases (“instability”) in the number of repeats in microsatellites throughout the genome (85). MMR proteins including MLH1, PMS2, MSH2 and MSH6 can form heterodimers to correct these mismatches. Mutations in any of these MMR genes, as well as in EpCAM (which leads to loss of MSH2 expression), can cause truncated protein products that result in tumorigenesis (86). These un repaired alterations contribute to carcinogenesis along a distinct pathway (the MSI pathway) that differs from the chromosomal instability pathway. About 15% colorectal cancers harbor MMR defects (deficient MMR, dMMR) (87,88). Among these cases, about 20% are caused by Lynch syndrome which include germline mutations in MMR genes (MLH1, MSH2, MSH6, and PMS2) while the majority of cases (80%) are caused by sporadic hypermethylation of the MLH1 gene promoter (86-91).

Deficient MMR results in the incorporation of mismatched nucleotides, with MMR-deficient colorectal cancer cells exhibiting 10–100 times the number of somatic mutations as those with proficient MMR (92-94). MSI-high status in colorectal cancer is associated with improved stage-specific prognosis although its predictive value is controversial (95-99). It has been observed that frameshift-induced neoepitopes in patients with MSI-high colorectal cancer are effectively recognized by the immune system. The better outcome of these patients is partly related to the high infiltration of activated CD8+ cytotoxic lymphocytes (CTL) and T helper 1 cells (Th1) (19,100-102).

MSI occurs in 10–39% of gastro-esophageal cancers. MSI in GC is associated with old age at onset, antral location, differentiated type, distinct mucinous or medullary histological patterns and reduced lymph node metastasis (103,104). In the largest studies, which were from Korea and examined five MSI markers, ~9% of GCs were MSI-H (MSI detected by more than one marker) (105,106). In GC patients from Europe, MSI was detected in 8–16% of cases (107,108). MSI-high GCs have been found to be associated with a denser accumulation of TILs and significantly associated with improved patient survival compared with MSI-stable/-low tumors (106).

MSI-high gastric tumors appear to have a significantly higher somatic mutation load (hypermutated). The most comprehensive molecular characterization of gastric AC was performed by the Cancer Genome Atlas (TCGA) and
examined 295 paired gastric AC cases using six molecular platforms (array-based somatic copy number analysis, whole-genome sequencing, array-based DNA methylation profiling, mRNA sequencing, microRNA sequencing and reverse-phase protein array). A new molecular classification describing four subtypes was proposed: MSI, EBV, genomically stable, chromosomal instable (CIN). Data from the MSI and EBV subgroups appeared to have greatest relevance for immune checkpoint treatment. MSI tumors appeared to have a significantly higher somatic mutation load (hypermutated). Genes mutated within MSI tumors revealed common alterations in major histocompatibility complex class I genes, including Beta2 microglobulin (B2M) and HLA-A. B2M mutations in colorectal cancers and melanoma result in loss of expression of HLA class 1 complexes (109), suggesting these events benefit hypermutated tumors by reducing antigen presentation to the immune system. In addition, elevated mutations were found in genes encoding targetable oncogenic signaling proteins. EBV tumors were enriched with PIK3CA mutations and extreme DNA hypermethylation. Interestingly, EBV tumors also showed amplification and/or expression of PD-L1 and PD-L2, providing rationale for testing immune checkpoint inhibitors in EBV-positive GC. Nivolumab monotherapy is currently being evaluated in a study of virus-associated tumors, including gastro-esophageal cancer (CheckMate358; NCT02488759).

MSI was detected in 6.6% (5/76) of patients with Barrett's-associated EAC, with the presence of MSI limited to AC cells, and not in adjacent Barrett's or normal epithelia (37,38). MSI has been less well studied in ESCC, although it has been detected in a subset of patients (110-112). Differences in the precise definition of MSI/MMR and test methods may account for some of these reported incidence differences (108).

**Predictive value of MSI in PD-1/PD-L1 blockade**

A direct correlation between MMR deficiency and an improved efficacy of PD-1/PD-L1 inhibition was recently suggested in a small study examining patients with advanced colorectal cancer (CRC) and non-CRC gastrointestinal tumors treated with pembrolizumab monotherapy. MSI was tested using the Promega system. The immune-related objective response rate (RR) was 40% (4/10) in MMR-deficient CRCs, as compared to 0% (0/18) in MMR-proficient CRCs. Patients with MMR-deficient non-CRC had responses similar to those of patients with MMR-deficient CRC [immune-related objective RR, 71% (5 of 7 patients)]. The median PFS and OS were not reached in the cohort with MMR-deficient CRC but were 2.2 and 5.0 months, respectively, in the cohort with MMR-proficient CRC (HR for disease progression or death =0.10; P<0.001; and HR for death, 0.22; P=0.05). Whole-exome sequencing revealed a mean of 1,782 somatic mutations per tumor in MMR-deficient tumors, as compared with 73 in MMR-proficient tumors (P=0.007), and high somatic mutation loads were significantly associated with prolonged PFS. A mean of 578 potential mutation-associated neoantigens were detected form dMMR tumors while only 21 such neoantigens were detected in MMR-proficient tumors. CD8 and PD-L1 IHC staining was not significantly associated with PFS or OS (94). Interestingly, in the cohort with dMMR non-CRC, all patients with sporadic tumors (n=6) responded to treatment, whereas only 3 of 11 patients with Lynch syndrome responded (94).

A subsequent report by the same group focused attention on dMMR non-CRC patients (66). Twenty-one patients with MMR-deficient tumors were given pembrolizumab (10 mg/kg intravenously every 2 weeks); the primary tumor was reported for 17 patients: ampullary (N=4), pancreas (N=4), biliary (N=3), small bowel (N=3), and gastric (N=3) cancers. For the ten evaluable patients whose results were reported at the 2016 ASCO GI meeting, ORR was 50% (5/10), disease control rate was 70% (7/10), and median OS was 21 months. After a median follow up of 7.6 months, the median duration of response (DOR) (range, 5.5 to 17+ months) and PFS were not reached. Further study of this agent in an expanded cohort is ongoing.

These preliminary data raise the hypothesis that MMR deficiency and somatic mutation burden in gastro-esophageal and other cancers could be a potential marker to predict efficacy from PD-1/PD-L1 treatment. As detected by whole genome or whole exome sequencing, gastric or esophageal cancers have among the highest somatic mutation rates among malignancies, with a median >4 mutations per megabase (MB) observed in each tumor type (113,114). As comparison, melanoma and squamous lung cancers have been reported to have the highest mutational load (10 or more mutations per MB), and pancreatic cancers have a lower rate (<1 mutation per MB).

**Table 4** shows studies in the gastro-esophageal cancers are currently ongoing by using compounds target the PD-1 pathway by either antagonizing PD-1 (pembrolizumab, nivolumab) or PD-L1 (atezolizumab).

Preliminary data in other cancers suggest that
tumors with high rates of somatic mutations (i.e., sun-exposed cutaneous melanoma, NSCLC in smokers, and microsatellite unstable colorectal carcinomas) have a higher chance of benefiting from immune checkpoint blockade than tumors with lower rates of somatic mutations (94,115,116). Exome sequencing in NSCLC patients treated with pembrolizumab showed that significantly elevated nonsynonymous mutation burden was strongly associated with clinical benefit from pembrolizumab treatment. In this study, the confirmed tumor RR (63% vs. 0%) and median PFS (14.5 vs. 3.7 months) was higher in patients (n=16) with a high vs. low nonsynonymous burden (with the cut-off at the median of 209 mutations per tumor). The PFS finding was confirmed in a validation cohort (n=18). In addition, the quantity of predicted neoantigens per tumor (a measure of immunogenicity) was found to correlate with mutation burden, and a high candidate neoantigen burden was associated with improved PFS. The molecular smoking signature, but not the patient self-reported smoking history, was correlated with treatment efficacy. In this trial, most patients had some degree of PD-L1 expression, limiting the ability to determine associations between mutation burden and PD-L1 expression. However, it appeared that mutation burden was able to distinguish patients who did and did not experience durable clinical benefit (defined as response lasting >6 months), even among those who had some degree of PD-L1 expression. Although this study had a limited sample size, these preliminary data raise the possibility that nonsynonymous mutation burden could serve as a potential biomarker for PD-1 blockade that can provide further predictive information beyond PD-L1 expression (116).

Future areas: checkpoint inhibition combined with chemo(radiotherapy)

Given that cancer cells evade the human immune system using different mechanisms, it is rational to consider combining PD-1/PD-L1 blockade with other therapies. Table 4 shows a sample of ongoing trials examining combination therapy.

Table 4

<table>
<thead>
<tr>
<th>Trial Name</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>NCT02443324</td>
<td>The combination of nivolumab with ramucirumab in locally advanced unresectable or metastatic gastric/GEJ cancer to evaluate the combination of nivolumab with ramucirumab.</td>
</tr>
</tbody>
</table>

With conventional chemotherapy and/or irradiation in curative settings

The possibility that irradiation and/or chemotherapy can induce neoantigen presentation and upregulate PD-L1 expression has been previously described (122-125). The combination of PD-1/PD-L1 blockade with chemotherapy or radiation treatment to enhance immune response has shown early promising results in animal models, and trials examining these combinations in human cancer are currently underway (NCT02268825, NCT02642809). At our institution, we will soon open a non-randomized trial in locally advanced resectable GEJ AC studying the combination of neoadjuvant pembrolizumab with concurrent carboplatin/paclitaxel-based chemoradiation followed by surgery and adjuvant pembrolizumab (NCT02730546).
Acknowledgements

None.

Footnote

Conflicts of Interest: The authors have no conflicts of interest to declare.

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